IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of

Ann Isabel SANZ MOUNERO

Atty. Ref.:

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Serial No.

10/537,897

Group:

1638

Filed:

June 7, 2005

Examiner:

Baum

For:

PLANTS HAVING MODIFIED GROWTH

CHARACTERISTICS AND A METHOD FOR MAKING

THE SAME

Commissioner for Paterits P.O. Box 1450 Alexandria, VA 22313-1450

Sir:

RULE 132 DECLARATION

- I, Ana Isabel Sanz Molinero, do hereby declare and say as follows:
- 1. I have reviewed the above-identified application, including the claims. I have also considered the documents listed herein. I am the inventor of the subject matter claimed in the above-identified application.
 - 2. I copy of my Curriculum Vitae is attached.
- 3. I am familiar with the evidence submitted to the U.S. Patent Office in the above on November 18, 2008. I have been advised by the assignee's U.S. representative that the U.S. Patent Office representative examining the above has invited the applicants to present the evidence in the form of a Declaration. The present paper is being filed in response to this invitation.
- 4. I have produced the following evidence or had the evidence produced under my direction.

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5. Example A - AtSTZparalog1: SEQ ID NO: 26 under the control of constitutive promoter GOS2

A DNA fragment encoding a 2XC2H2 protein represented in the application as filled by SEQ ID NO: 27 was isolated from an *Arabidopsis thaliana* seedling cDNA library (Invitrogen, Paisley, UK) by PCR amplification and subsequent cloning in an entry clone vector according to the methods described in Example 1 of the present application.

SEQ ID NO: 27 polypertide was encoded by the longest open reading frame of SEQ ID NO: 26 (AtSTZparalog1).

The primers used for the PCR amplification were as follows:

Forward primer: Ggggaca agtttgtacaaaaaagcaggcttaaacaatggccctcgaagcg Reverse primer: Ggggaccactttgtacaagaaagctgggttcgagtattagatttttaaagataaatc

The entry clone was subsequently used in an LR reaction with a destination vector used for rice transformation to generate the plant expression vector pGOS2::AtSTZparalog1. The constitutive promoter, GOS2, was mentioned in Table 10 on page 48 of the application as filed as being a promoter useful in the methods of the invention.

Phenotypic characterization of the transformed plants was carried out essentially as described in Example 3 of the present application. The results are shown in Table I below.

Table I: Results of phenotypic characterization of T2 rice plants transformed with pGOS2::AtSTZparalog1

pGOS2::AtSTZparalog1	
I ^p arameter	% increase in the transgenic plants compared to the nullyzygous plants



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, ;		
pGOS2::AtSTZparalog1		
Parameter	% increase in the transgenic plants compared to the nullyzygous plants	
boveground	The state of the s	
area	10	
Root Area lotal Seed	4	
Weight	48	
lumber of filled seeds	46	
lotal number of seeds	12	
Seed filing rate	30	
Flowers per Fanicie	8	
l arvest index	35	
Days to flowering	4	

The above results show that overexpression of the nucleic acid represented by SEQ ID NO: 27 (encoding the 2XC 2H2 zinc finger protein represented by SEQ ID NO 26) under the control of a constitutive promoter (GOS2) gives:

Increased plant yield (in the form of increased root area, total seed weight, total number of seeds, number of filled seeds, seed filling rate, flowers per panicle and harvest index);

Increased leaf surface area (as manifested by increased aboveground area); and Prolonged vegetative growth (where the time to flower was on average 4% longer in transgenic plants compared to corresponding nullyzygous plants).

6. Example B - A STZparalog1: SEQ ID NO: 26 under the control of seed-specific promoter prolamin

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A DNA fragment encoding a 2XC2H2 protein represented by SEQ ID NO: 27 was isolated from an *Arabid posis thaliana* seedling cDNA library (Invitrogen, Paisley, UK) by PCR amplification and subsequent cloning in an entry clone vector according to the methods described in Example 1 of the present application. SEQ ID NO: 27 polypeptide was encoded by the longest open reading frame of SEQ ID NO: 26 (AtSTZparalog1).

The primers used for the PCR amplification were as follows:

Forward primer: Ggggaca agtttgtacaaaaaagcaggcttaaacaatggccclcgaagcg Reverse primer: Ggggaccactttgtacaagaaagctgggttcgagtattagatttttaaagataaatc

The entry clone was subsequently used in an LR reaction with a destination vectors used for rice transformation to generate the plant expression vector pPROLAMIN::AtSTZparalog1. The seed-specific promoter, prolamin, was mentioned in Table 10 on page 48 of the application as filed as being a promoter useful in the methods of the invention.

Phenotypic characterization of the transformed plants was carried out essentially as described in Example 3 of the present application. The results are shown in Table II below.

Table II: Results of phenotypic characterization of T2 rice plants transformed with pPROLAMIN::AtSTZparalog1.

pPR(pPROLAMIN::AtSTZparalog1		
Parameter	% increase in the transgenic plants compared to the nullyzygous plants		
Apoveground			
area	4		
T∳tal Seed			
<u>W</u> eight	8		

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pPROLAMIN::AtSTZparalog1		
Parameter	% increase in the transgenic plants compared to the nullyzygous plants	
Number of filled seeds	7	
otal number of seeds	5	
Flowers por panicle	4	
larvest index	5	

The above results show that overexpression of the nucleic acid represented by SEQ ID NO: 27 (encoding the 2XC2H2 zinc finger protein represented by SEQ ID NO 26) under the control of a seed-specific promoter (protamin) gives:

Increased plant yield (in the form of increased aboveground area, total seed weight, total number of seeds, number of filled seeds, flowers per panicle and harvest index); and

Increased leaf sunface area (as manifested by increased aboveground area).

7. Example C - OsSTZ(ortholog): SEQ ID NO: 36 under the control of root-specific promoter RCc3

A DNA fragment comprising the coding region of SEQ ID NO 36 was PCR amplified and cloned using methods essentially as described in the Examples section of the present application.

The primers used for the PCR amplification were as follows:

Forward primer: ggggaccaptttgtacaaaaaagcaggcttaaacaatgtcgagcgcgtcgt Reverse primer: ggggaccaptttgtacaagaaagctgggtctgaattacgcggtgagaag



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A plant transformation vector carrying the coding region of SEQ ID NO 36 under the control of the root specific promoter, RCc3, was made, giving construct CD10315 described in Table 9 on page 47 of the present application. Agrobaterium-mediated transformation of rice plants was carried out to generate transgenic rice plants carrying the construct CD10315.

The results of the phenotypic evaluation of the CD10315-transgenic plants of the T2 generation are shown in Table III below.

Table III: Results of phenotypic characterization of T2 rice plants transformed with pRCc3::OsSTZorthblog.

			
	CD10315 plants		
Parameter	% increase in the transgenic plants compared to the nullyzygous plants		
Aboveground			
aļea	6		
Root Area	5		
Total Seed Weight	29		
Number of filled seeds	29		
Seed filling	16		
Total number of seeds	17		
Harvest index	25		

The above results show that overexpression of a nucleic acid encoding the 2XC2H2 zinc finger protein represented by SEQ ID NO 36) under the control of a root-specific promoter (RCc3 gives:

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Increased plant yield (in the form of increased aboveground area, root area, total seed weight, total number of seeds, number of filled seeds, seed filling rate and harvest index); and

Increased leaf surface area (as manifested by increased aboveground area).

8. Example D - AtSTZparalog2: SEQ ID NO:28 under the control of the betaexpansine EXPB9 (PRØ061 2)





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A DNA fragment encoding a 2XC2H2 protein represented in the application as filed by SEQ ID NO: 29 was isolated from an *Arabidopsis thaliana* specifing cDNA library (Invitrogen, Paistey, UK) by PCR amplification and subsequent cloning in an entry clone vector according to the methods described in Example 1 of the present application. SEQ ID NO: 29 polypoptide was encoded by the longest does reading frame of SEQ ID NO: 28 (AtSTZparalog2).

The primers used for the PCR amplification were as follows:

Forward primer:

qqqqacaagtttgtacaa**aaaa**gcaggottaaacaatggcacttgaaactdtcact

Reverse primer:

ggggacoactttptacaagaaagctgggtttcctaggtttatgtttaggg

The entry clone was subdequently used in an LR reaction with a destination vector used for rice transformation to generate the plant expression vector pEXPB9::AtSTZparalog2. The PCR amplified DNA fragmented was operably linked to the promoter PROO61_2 (pEXPB9: beta-expansine EXPB9), which is described in Table 10 in Table 10 on page 48 of the application as filled

Phenotypic characterization of the transformed plants was carried out essentially as described in Example 3 of the present application. The results are shown in Table I below.

Table I: Results of phenotypic characterization of T1 rice plants transformed with pEXPB9::AtSTZparalog2

	pEXPB9::AtSTZparalog2	
Paramete		% increase in the transgenic plants compared to the nullyzygous plants
-	f filled seeds	6.2
Seed filin	j rato	3.4

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Harvest Index	y. y
i	1
from the same of t	125 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
Number of panicles	3.8
The second secon	P. C
	1.

The above results show that overexpression of the nucleic acid represented by SEQ ID NO: 28 (encoding the 2XC2H2 \$inc finger protein represented by SEQ ID NO 29) under the control of a promoter expressed in young expanding tissues (pEXP9) gives an increase in plant yield a shown by the increased in the number of filled seeds, the increase in the seed filling rate (proportion of filled seed with respect to the total number of seeds harvested per plant, increased in the harvest index and the increase in the number of panicles per plant.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these staterhents were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Signed this 20 day of //a , 2009.

(print name) Ana Isabel Sanz Molinero